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EDWARDS ANGELI, PALMER & DODGE LLP P.O. BOX 55874 BOSTON, MA 02205			NEGIN, RUSSELL, SCOTT	
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary	Application No. 10/698,599	Applicant(s) WIGSTROM ET AL.
	Examiner RUSSELL S. NEGIN	Art Unit 1631

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --
Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If no period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED. (35 U.S.C. § 133).

Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

1) Responsive to communication(s) filed on 02 September 2009.
 2a) This action is FINAL. 2b) This action is non-final.
 3) Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

4) Claim(s) 1-3,7-34 and 36-189 is/are pending in the application.
 4a) Of the above claim(s) 1 is/are withdrawn from consideration.
 5) Claim(s) _____ is/are allowed.
 6) Claim(s) 1-3,8-34 and 36-189 is/are rejected.
 7) Claim(s) _____ is/are objected to.
 8) Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

9) The specification is objected to by the Examiner.
 10) The drawing(s) filed on _____ is/are: a) accepted or b) objected to by the Examiner.
 Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
 Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
 11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
 a) All b) Some * c) None of:
 1. Certified copies of the priority documents have been received.
 2. Certified copies of the priority documents have been received in Application No. _____.
 3. Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

1) Notice of References Cited (PTO-892)
 2) Notice of Draftsperson's Patent Drawing Review (PTO-948)
 3) Information Disclosure Statement(s) (PTO/SB/06)
 Paper No(s)/Mail Date _____

4) Interview Summary (PTO-413)
 Paper No(s)/Mail Date _____

5) Notice of Informal Patent Application
 6) Other: _____

DETAILED ACTION

Continued Examination Under 37 CFR 1.114

A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on 2 September 2009 has been entered.

Comments

Claim 7 is withdrawn from further consideration pursuant to 37 CFR 1.142(b) as being drawn to a nonelected Species, there being no allowable generic or linking claim. Election was made **without** traverse in the reply filed on 10 April 2008.

Claims 1-3, 7-34, and 36-189 are pending in the instant Office action.

Claims 1-3, 8-34, and 36-189 are examined in the instant Office action.

It is noted that claim 7 does not comply with 37 CFR 1.121 because it has the incorrect status identifier; instead of "Previously presented," claim 7 should be "Withdrawn." However, in the interest of advancing prosecution, the application is examined as it stands.

Withdrawn Objections/Rejections

The objection to claim 166 is withdrawn in view of amendments filed to the instant claim on 2 September 2009.

The rejections of claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124-134, 137-139, 142-149, 151, 153-155, and 159-161 under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of Agilent [Agilent capillary electrophoresis system; brochure; 12 pages; published 1 September 2001] are withdrawn in view of amendments filed to the instant set of claims on 2 September 2009.

The rejections of claims 14, 19-23, 99, and 156 under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure as applied above, and further in view of Colton et al. [Electrophoresis, 1998, volume 19, pages 367-382] are withdrawn in view of amendments filed to the instant set of claims on 2 September 2009.

The rejections of claims 34, 40-42, 150, and 152 under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure as applied above, and further in view of Katayama et al. [Analytical Chemistry, 1998, volume 70, pages 2254-2260] are withdrawn in view of amendments filed to the instant set of claims on 2 September 2009.

The rejections of claims 95-96, 123, 135-136, 140-141, and 157-158 under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure as applied above, and further in view of Jardemark et al. [Analytical Chemistry, 1997,

volume 69, pages 3427-3434] are withdrawn in view of amendments filed to the instant set of claims on 2 September 2009.

The rejection of claim 125 under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure as applied above, and further in view of Couderc et al. [Electrophoresis, 1998, volume 19, pages 2777-2790] is withdrawn in view of amendments filed to the instant claim on 2 September 2009.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

The following rejections are newly applied and are necessitated by amendment:

35 U.S.C 103 Rejection #1:

Claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124, 126-134, 138-139, 142-149, 151, 153-155, and 159-161 are rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of Agilent [Agilent capillary electrophoresis system; brochure; 12 pages; published 1 September 2001] in view of Ishmagilov et al. [WO 02/086333; published 31 October 2002; filed 24 April 2002; benefit date 25 April 2001].

Claim 1 is drawn to a computer program product comprising a computer readable medium having computer readable program code embodied in the medium for causing an application program to execute on a computer, wherein the program product comprises instructions for controlling one or more functions of a microfluidic substrate in response to received data regarding one or more substrate properties.

Claim 2 is drawn to the same subject matter as instant claim 1 with the additional limitation of controlling one or more functions of a microfluidic substrate in response to received data regarding one or more properties of a sensor in fluid communication with at least one microchannel of the substrate.

Claims 1 and 2 are further limiting wherein the one or more function comprises scanning a cell based biosensor in electrical communication with an electrode relative to multiple substantially separate fluid streams from one or more outlets of one or more microchannels in the substrate by moving the sensor, moving the substrate, moving both the sensor and the substrate, and/or varying the pressure of one or more of the microchannels.

Claim 3 is drawn to a computer program product comprising a computer readable medium having computer readable program code embodied in the medium for causing an application program to execute on a computer, wherein the program product comprises instructions for controlling the scanning of a cell based biosensor to multiple substantially separate fluid streams from a plurality of outlets of more or microchannels in the substrate by varying the pressure of one or more of the microchannels.

Klein et al. teaches a multi-channel automated capillary electrophoresis analyzer. Specifically, Figure 8 and column 10, lines 22-45 of Klein et al. teach the computerized limitations behind the capillary electrophoresis system. Specifically, this section teaches computer program product comprising a computer readable medium causing operation of the computer (in the case of Klein et al. the computer controlling the CE apparatus is an IBM with a floppy disk). The substrate in this instance is the capillary itself, and the properties being measured relate to flow of liquid through the capillary (see Figure 3 of Klein et al.).

However, Klein et al. does not teach scanning of sensors or instructions for controlling the scanning of sensors. Additionally, Klein et al. does not teach substantially separate fluid streams.

Agilent is a brochure describing the benefits of using an Agilent capillary electrophoresis system for measuring biomolecules. Specifically, the fourth page of the brochure measures the migration time of an oligonucleotide sensor. The brochure itself gives instructions for detecting sensors using the CE apparatus in the form of

specifications on the penultimate page of the brochure. Pressure variance is illustrated on page eighth page of the brochure.

However, Klein et al. and Agilent do not teach scanning of a cell based biosensor to multiple SUBSTANTIALLY SEPARATE fluid streams. Applicant defines substantially separate fluid streams on page 19, lines 20-21 of the specification as collimated laminar flow.

The document of Ishmagilov et al. studies fluidic switches and methods for controlling flow in microfluidic systems.

Lines 22-24 of page 1 of Ishmagilov et al. teaches that flow lines of laminar flow (as opposed to turbulent flow) are in parallel (i.e. they are collimated). Figure 10 of Ishmagilov et al. illustrates this property of collimated laminar flow in simple geometries.

With regards to claims 54-57, 60-61, 67-69, 73, 126, and 161 Figure 8 of Klein et al. illustrates a GUI in the form of a monitor that regulates and displays results of the CE in response to user commands and acquires, retrieves, and manages data from the CE apparatus. Additionally, Figure 8 is viewed as a microfluidics workstation linked to a microfluidic substrate (i.e. the capillaries). Figure 8 also illustrates a memory board.

With regard to claims 103-104, the abstract of Klein et al. indicates the plurality of capillary substrates. The Agilent brochure indicates, as discussed above, movement of the sensor through the capillary. When buffer is delivered to each of the channels (as in

Klein et al.), there is buffer delivered to each adjacent capillary (i.e. capillaries with sensors/agents).

With regard to claims 8, 15-16, and 87-88, Klein et al. illustrates movement of an aqueous liquid from a reservoir through the capillary into a second reservoir in Figure 3.

With regard to claims 17 and 89, Klein et al. is a capillary electrophoresis procedure that applies electric fields to capillaries [see abstract of Klein et al.]

With regard to claims 18 and 38, Klein et al. teaches the computerized limitations governing the electrophoresis in Figure 8 and column 10, lines 22-45 of Klein et al.

With regard to claims 24-25, 48-49, 51, 93, 100-101, and 105, Figure 6 of Klein et al. illustrates automated delivery of four fluids to the capillaries, including flow of buffer. The buffer includes ionic NaOH. This buffer is a component of the filled capillary.

With regard to claims 26, 52, 65, and 76, the fourth page of the Agilent brochure indicates the movement of oligonucleotide sensors and buffer through the capillary substrate in a continuous manner.

With regard to claims 27, 110, 114-117 and 121, the abstract of Klein et al. indicates use of a vacuum to regulate pressure and help move liquid in the capillary channel.

With regard to claims 9-11 and 53, Figure 3 of Klein et al. illustrates a UV detector that scans near the output of the capillary substrate. The Agilent brochure indicates, as discussed above, movement of the sensor through the capillary. The figure on the fourth page of the Agilent brochure illustrates signal data.

With regard to claims 12-13, the fifth page of the Agilent brochure indicates the physiological response of the production of DNA and protein resulting from PCR hybridization reaction with PCR agents.

With regard to claims 28, 32-33 and 36-37, instructions for delivering agent/buffer to the capillary system are illustrated in the monitor on the second page of the Agilent brochure. The computer has a memory. The figure on the fourth page of the Agilent brochure illustrates response data.

With regard to claims 29-30, the abstract of Klein et al. teaches a plurality of capillaries and the fourth page of the Agilent brochure illustrates the result of delivery of an agent to a capillary substrate.

With regard to claim 31, the fourth page of the Agilent brochure indicates a change in amount of oligonucleotide added to the capillary substrate (i.e. the intensities of each electropherogram differ depending on whether aqueous or organic solvent is used with the oligonucleotides).

With regard to claims 39, 50, 109 and 120, Klein et al. teaches modifications that cause temperature of the capillary to be adjusted in column 15, lines 46-55.

With regard to claims 43-47, 107-108 and 111-113, Figure 8 of Klein et al. teaches a processor in communication with a macroscale device that is in communication with a microfluidic device. Figure 8 also illustrates a power supply. This apparatus performs capillary electrophoresis using a voltage and a current to the substrate (i.e. a capillary channel).

With regard to claim 58, the third page of the Agilent brochure indicates a linearity in the scanning process for capillary electrophoresis.

With regard to claim 59, the second page of the Agilent brochure indicates the presence of the sensor in the capillary.

With regard to claims 62-64, the fluids/sensor in the capillary substrate are scanned continuously at the detector window near the output of the capillary and the time of the sensor migration is recorded in the fourth page of the Agilent brochure. Consequently, the recorded property is the migration time.

With regard to claim 66, electropherograms of proteins are illustrated in the fifth page of the Agilent brochure.

With regard to claims 70-72 and 85, column 10, lines 58-64 of Klein et al. teaches use of an electrical current monitoring resistor to monitor and regulate current and voltage across the capillary substrate in response to feedback or a user or data acquisition system.

With regard to claim 74, the workstation illustrated on the second page of the brochure illustrates the instructions for performing the capillary electrophoresis.

With regard to claims 75 and 77, the ninth page of the Agilent brochure illustrates the CE-MS procedure with the MS abundances mapped to a database of biomolecular identifiers in the upper left corner of the figure on this ninth page with electropherograms.

With regard to claims 78-80, 90-92 and 131, the apparatus in Klein et al. is a macroscale device regulated by a power supply as illustrated in Figure 8 of Klein et al. and receives instructions from the computer. Furthermore, the electrical power supply illustrated in Figure 8 is regulated by the computer system to generate the desired electric field across the capillary.

With regard to claims 81-84 and 86, the second and third pages of the Agilent brochure illustrate a macroscale device comprising a detector for detecting response to a sensor in the form of signal data. The fourth page of the Agilent brochure illustrates a physiological response to oligonucleotides in which a mathematic operation is performed on the data after acquisition to generate the electropherogram.

With regard to claims 94 and 97-98, the electrophoresis apparatus in the third page of the Agilent brochure illustrates equipment in which an electric field applied to capillary substrate containing a sensor. Specifically, the monitor on the third page of the Agilent brochure allows input data regarding the agent (i.e. amount of agent to be used).

With regard to claims 118-119 and 122, the fifth page of the Agilent brochure illustrates the separation of a group of proteins wherein the capillary substrate comprises the protein sensor.

With regard to claim 102, the abstract of Klein et al. teaches a plurality of substrates (i.e. capillaries) in the apparatus.

With regard to claim 124, Figure 3 of Klein et al. illustrates use of a UV light source to monitor the contents of the capillary.

With regard to claim 127-130 and 138, the computer on the third page of the Agilent brochure illustrates a graphical user interface with a substrate and apparatus properties and function parameters which include application of the electric field or vacuum. Additionally, a representation of a substrate is shown in the third page of the Agilent brochure.

With regard to claims 132-133, the sensor in the CE system comprises a memory (see third page of Agilent brochure and Figures 3 and 8 of Klein et al.) recording the scanning data across fluid streams at a sensor location and sensor response as a function of time.

With regard to claim 134, the cover Figure of Klein et al. illustrates a circular stage for receiving substrate molecules that is rotatable in a circular (i.e. x and y directions).

With regard to claim 139, the fourth page of the Agilent brochure illustrates a selection of and an operation of the abundance as a function of migration time of the sensor in the capillary under an applied electrical field.

Claim 142 is further limiting wherein the system further comprises a first computer program product according to any of claims 1-3, a second computer program product comprising computer program code from acquiring data relating to properties of a sensor in fluid communication with at least one channel of the microfluidic substrate; and a data processing system for accessing the data relating to properties of the sensor and for providing the data to the first computer program product.

Claim 143 is further limiting wherein the instructions executed by the first program product affects the second program product.

The patent of Klein et al. documents the first computer system in Figure 8 and column 10, lines 20-45. The Agilent brochure details a second computer system used to detect the presence of a sensor in the capillary substrate channel.

With regard to claim 144, Klein et al. teaches use of a vacuum to regulate pressure in the abstract.

Claim 145 is further limiting wherein a sensor in fluid communication with at least one microchannel on a microfluidic substrate;

--providing data to a computer program product according to any of claims 1-3, wherein in response to the data provided, the computer program product provides instructions to a scanning mechanism to execute one or more scanning functions such that the substrate, the sensor, or the substrate and the sensor move relative to one another, and/or such that pressure is altered in at least one microchannel of the substrate.

The fourth page of the Agilent brochure indicates the movement of oligonucleotide sensors and buffer through the capillary substrate in a continuous manner. The concentration of sensor varies with time as is measured when scanned by a UV detector proximate to the output of the capillary (see Figure 3 of Klein et al.). Additionally, Klein et al. describes a vacuum to regulate pressure in the capillary in the abstract.

With regard to claims 146 and 151, the capillary system in Agilent brochure (i.e. fourth page) indicates a fluid stream (i.e. buffer) with a sensor/agent (i.e. oligonucleotide) as a function of time.

With regard to claim 147, Klein et al. discloses an electrophoresis system with a plurality of capillary substrates. The Agilent brochure introduces sensors into the capillaries.

With regard to claims 148-149, Figure 6 of Klein et al. illustrates a pre-programmed set of four chemicals automatically inserted into the apparatus. The scans of sensors as a function of migration time are continuous and illustrated in the fourth page of the Agilent brochure.

With regard to claim 153-155 and 159, Figure 3 of Klein et al. and the abstract show the varying of pressure and the UV detection of the sensor (i.e. such as those disclosed in Agilent brochure) near the output of the capillary. The sensor is detected in response to UV light, and the flow of the surrounding fluid in the channel. The detection of the sensor is measured in response to an electrical field.

With regard to claim 160, the third page of the Agilent brochure illustrates a graphical user interface where there is data that is entered and the sensor is then scanned for optical properties.

It would have been to someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al. by use of the sensors, agents, and capillary electrophoresis apparatus of the Agilent brochure wherein the motivation would have been that the use of a sensor gives the apparatus an entity with which to measure migration time [i.e. the Figure on the fourth page of Agilent]. Furthermore, the computer GUIs in Agilent provide a further means of automation to the apparatus of Klein et al. Furthermore, the multiple capillaries in the

apparatus with the computer system of Klein et al. enable multiple experiments (i.e. of sensors such as those in computer system/apparatus of Agilent) to be performed at once.

It would have been further obvious to modify the multi-capillary electrophoresis system of Klein et al. and the sensors, agents, and apparatus of the Agilent brochure by use of the collimated and substantially separate laminar flows of Ishmagilov et al. wherein the motivation would have been that the laminar flows of the system of Ishmagilov et al. allows adjacent and parallel flows of different solutions without barriers between the solutions and without mixing of the different channels of flow [see, for example, Figure 10 of Ishmagilov et al. and the Background section of Ishmagilov et al.]

There would have been a reasonable expectation of success in combining Klein et al., the Agilent brochure, and Ishmagilov et al. because each study analogously pertains to flows of sensors and agents through microchannels.

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant states on page 32 that the amended version of claims 1-3 recites:

...scanning a cell based biosensor in electrical communication with an electrode relative to multiple substantially separate fluid streams from one or more outlets...

Applicant argues that Klein et al. and the Agilent brochure do not teach this amended limitation.

Specifically, applicant first reiterates the meaning of "scanning of a sensor relative to one or more channels in a microfluidic substrate" which is stated again below [paragraph bridging pages 20-21 of the specification]:

...refers to exposure of the sensor to a plurality of fluid streams from at least one channel in the substrate. This **MAY BE** achieved by moving a sensor past one or more channel outlets in a stationary substrate providing such streams or by moving the substrate relative to a stationary sensor so that it is exposed to streams from one or more channel outlets of the substrate. Scanning **MAY ALSO BE** achieved by moving both the substrate and the sensor. Exposure to a plurality of fluid streams from a single channel **MAY BE** achieved by providing different fluid streams... ...from the single channel and/or by intermittently stopping the flow of fluid from an outlet of the channel in proximity to the sensor. In an embodiment where the sensor is stationary, scanning **CAN BE** done by varying pressure at one or more channels. Combinations of the above scanning mechanisms **MAY BE** used during a scanning process and variations of such combinations are obvious and encompassed within the scope of the invention.

In the above passage, the only part of the definition that is REQUIRED is that the sensor be exposed to a plurality of fluid streams from at least one channel in the substrate; the remaining embodiments are exemplary. Furthermore, the invention of Klein et al. and the brochure of Agilent teach this definition of "scanning a sensor" in that the sensors are applied to the plurality of channels (i.e. capillaries- each with its own fluid stream) of Klein et al.

With regard to the limitation requiring "substantially separate fluid streams," (which is defined in the specification to be collimated laminar flow), the document of Ishmagilov et al. teaches this limitation in Figure 10.

The following rejections are newly applied and are necessitated by amendment:

35 U.S.C. 103 Rejection #2:

Claims 14, 19-23, 99, and 156 are rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure in view of Ishmagilov et al. as applied to claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124, 126-134, 138-139, 142-149, 151, 153-155, and 159-161 above, and further in view of Colton et al. [Electrophoresis, 1998, volume 19, pages 367-382].

Claims 14, 19-20, 22 and 99 are further limiting comprising delivering an agent to a sensor.

Claim 21 is further limiting wherein a parameter of the agent comprises a property of the agent.

Claim 23 is further limiting wherein the data is dose-respondent.

Claim 156 is further limiting wherein the result is a different electrical charge for the sensor.

Klein et al., Agilent, and Ishmagilov et al. make obvious a capillary electrophoresis system for detecting the presence and properties of sensors and analytes, as discussed above.

Klein et al., Agilent, and Ishmagilov et al. do not describe interactions between the sensor and analyte in the capillary substrate.

The review of Colton et al. studies the technique of affinity capillary electrophoresis, which as illustrated in Figure 1 on page 369 shows how capillary electrophoresis is able to measure the binding constant between a receptor (i.e. sensor)

and a ligand (i.e. agent) by comparing the migration times of the receptor, ligand, and receptor-ligand complex. Figure 1 also illustrates that an electrical property (i.e. the charge) changes as a result of complex formation.

It would have been obvious someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al., Agilent, and Ishmagilov et al. by use of the affinity capillary electrophoresis of Colton et al. wherein the motivation would have been that by combining binding affinities (i.e. of agents relative to sensors) with capillary electrophoresis, a more advanced means for determining binding constants of physiologically relevant processes is developed [see introduction on pages 367-368 of Colton et al.].

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant argues that since the first 35 U.S.C. 103 rejection is allegedly deficient, the instant rejection is also deficient. This argument is not persuasive because the combination of references teaches all of the limitations of the instantly rejected claims.

The following rejections are newly applied and are necessitated by amendment:

35 U.S.C. 103 Rejection #3:

Claims 34, 40-42, 150, and 152 are rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure in view of Ishmagilov et al.

as applied to claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124, 126-134, 138-139, 142-149, 151, 153-155, and 159-161 above, and further in view of Katayama et al. [Analytical Chemistry, 1998, volume 70, pages 2254-2260].

Claim 34 is further limiting wherein sensors are superfused at different time intervals according to instructions from the computer readable media.

Claim 40 is further limiting wherein the parameters are altered for the one or more functions in response to a measured condition of the microfluidic substrate or sensor.

Claim 41 is further limiting wherein the measured condition comprises an electrophoresis event.

Claim 42 is further limiting wherein the substrate function comprises time to complete scanning.

Claim 150 is further limiting wherein the sensor is paused during a given time interval.

Claim 152 is further limiting wherein the fluid streams provide interdigitating fluid streams of agent and buffer and the sensor is sequentially scanned across the fluid streams.

Klein et al., Agilent, and Ishmagilov et al. make obvious a capillary electrophoresis system for detecting the presence and properties of sensors and analytes, as discussed above.

Klein et al., Agilent, and Ishmagilov et al. do not teach superfusing, interdigitating, altering in response to a property of a sensor, or pausing during an electrophoresis run.

The study of Katayama et al. studies stable capillary coating with successive multiple ionic polymer layers. The "Procedure of SMIL coating" as taught in the paragraph bridging columns 1 and 2 of page 2255 includes pauses before each injection with buffered agents/sensors. Consequently, each layer is superfused at different time intervals (i.e. see Figure 1 on page 2255 of Katayama et al.). The electrophoresis event being modified by successively (i.e. interdigitating) coating the capillary with ionic layers is to change in direction of the electroosmotic flow (again, see Figure 1 of page 2255 of Katayama et al.). Time to complete scanning is shown in Figure 5 of Katayama et al.

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al. and Agilent and the microfluidic apparatus of Ishmagilov et al. by use of the SMIL capillary coating procedure of Katayama et al. wherein the motivation would have been that the modifications (i.e. interdigitating, pausing) would have allowed a more advanced, regulated electrophoresis process with less interaction between the sensor/agent and the wall of the capillary [see for example, introduction on page 2254 of Katayama et al.]

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant argues that since the first 35 U.S.C. 103 rejection is allegedly deficient, the instant rejection is also deficient. This argument is not persuasive because the combination of references teaches all of the limitations of the instantly rejected claims.

The following rejections are newly applied and are necessitated by amendment:

35 U.S.C. 103 Rejection #4:

Claims 95-96, 123, 135-137, 140-141, and 157-158 are rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure in view of Ishmagilov et al. as applied to claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124, 126-134, 138-139, 142-149, 151, 153-155, and 159-161 above, and further in view of Jardemark et al. [Analytical Chemistry, 1997, volume 69, pages 3427-3434].

Claim 95-96 are further limiting wherein the electric field can electroporate a membrane of the cell structure.

Claims 123 and 157 are further limiting wherein the sensor comprises a cell or cell fraction.

Claims 135, 140, and 158 are further limiting wherein the workstation or program suite comprises executing patch clamp analysis. Claim 141 is further limiting comprising computer code for analyzing patch clamp data.

Claims 136-137 are further limiting comprising mechanisms for controlling movement of the stage.

Klein et al., Agilent, and Ishmagilov et al. make obvious a capillary electrophoresis system (including computer code and computer readable media) for detecting the presence and properties of sensors and analytes, as discussed above.

Klein et al., Agilent, and Ishmagilov et al. do not teach use of cells and patch clamp analysis.

The study of Jardemark et al. studies patch clamp detection in capillary electrophoresis.

Specifically, Figure 1 on page 3429 of Jardemark et al. illustrates the patch clamp procedure with the cellular components undergoing electrophoresis (i.e. the sensor comprises fragments of the cells). The caption of Figure 1 indicates that the setup is capable of being manipulated with "micromanipulators."

With regard to claim 137, the mouse on the Agilent computer system is interpreted to be a joystick (i.e. see second and third pages of the Agilent Brochure).

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al. and Agilent and the microfluidic substrate of Ishmagilov et al. by use of the patch clamp

capillary electrophoresis apparatus of Jardemark et al. wherein the motivation would have been that this patch clamp method on cells is useful in fractionating biological tissues and can sense very low concentration and small-sized analytes (see penultimate paragraph of the introduction on page 3428 of Jardemark et al.). It would have been further obvious to apply the pressures (positive and negative) and waste collection of Klein et al. to the microfluidic device of Jardemark et al. wherein the motivation would have been that such pressures and waste collections permit more convenient analyses of the cell based biosensors [see Figure 1 and its caption in Jardemark et al.]

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant argues that since the first 35 U.S.C. 103 rejection is allegedly deficient, the instant rejection is also deficient. This argument is not persuasive because the combination of references teaches all of the limitations of the instantly rejected claims.

The following rejections are reiterated from the previous Office action:

35 U.S.C. 103 Rejection #5:

Claims 162-189 are rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure as applied to claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124, 126-134, 138-139, 142-149, 151, 153-155, and 159-

161 above, and further in view of Jardemark et al. [Analytical Chemistry, 1997, volume 69, pages 3427-3434].

Claims 162-189 are drawn to a computer program product and system that utilize a patch clamp system and pressure (negative and positive) to introduce cell based biosensors into a plurality of microchannels in communication with electrodes that then generate electric fields through these microchannels. Claims 162-189 comprise a plurality of reservoirs (i.e. chambers) used to hold the cell based biosensors before and after electrophoretic analysis. The cell based biosensors are SEQUENTIALLY exposed to a plurality of fluid streams from one or more microchannels.

Claim 162 is drawn to a computer program product comprising:

--a computer readable medium having computer readable program code embodied in the medium for causing an application program to execute on a computer, wherein the program product comprises instructions for controlling a fluid delivery control mechanism;

--delivers a cell based biosensor via a fluid through a first microchannel in a microfluidic substrate to a sensor chamber comprising one or more electrodes in communication with a sensor chamber which form a patch clamp system;

--applies a negative pressure to a second microchannel so as to position said cell based biosensor in electrical communication with the one or more electrodes;

--scanning the cell based biosensor to a plurality of fluid streams from one or more microchannels.

As discussed above, Klein et al. and Agilent teach a computer readable medium in a computer product causing controlled fluid delivery of a cell based biosensor (i.e. DNA) using a plurality of electrodes [i.e. see cover Figure of Klein et al.] Negative

pressure (i.e. a vacuum) to the microchannel initiates positioning of the DNA in the microchannel. Klein et al. describes this process for a plurality of capillaries in sequence [see Figure 2 of Klein et al.] Klein et al. and Agilent do not show a patch clamp system.

As discussed above, the study of Jardemark et al. studies patch clamp detection in capillary electrophoresis.

Specifically, Figure 1 on page 3429 of Jardemark et al. illustrates the patch clamp procedure with the cellular components undergoing electrophoresis (i.e. the sensor comprises fragments of the cells). The caption of Figure 1 indicates that the setup is capable of being manipulated with "micromanipulators."

With regard to claims 163-165, Figure 8 of Klein et al. illustrates computer program products for acquiring data from a plurality of microchannels in the electrophoresis apparatus. Additionally, Figures 6 and 8 of Klein et al. illustrate the data acquisition system operably linked to a microfluidic substrate.

With regard to claim 166, Figure 3 of Klein et al. illustrates a microfluidic substrate comprising a microchannel in communication with sensor chambers comprising electrodes.

With regard to claim 167, Figure 1 of Jardemark et al. illustrates an in chip patch clamp detection system for capillary electrophoresis.

With regard to claims 168-169, Figures 6 and 8 of Klein et al. illustrate a means for providing data to alter the substrate functions.

With regard to claim 170, Figure 3 of Klein et al. illustrates application of an electric field across an electrically conducting surface (i.e. a capillary) which contains the biosensor.

With regard to claims 171-172, the caption of Figure 1 of Jardemark et al. indicates that the patch-clamp setup is capable of being manipulated with "micromanipulators" capable of fragmenting cell membranes.

With regard to claims 173-174 and 177, Figures 3, 6, and 8 of Klein et al. illustrate a computer program product for using a computer to input data relating to the electric field properties of the system that comprises using the detector of Figure 3 of Klein et al.

With regard to claim 175-176, page 5 of Agilent illustrates the input to the detector regarding the product of a PCR (i.e. hybridization) reaction of DNA.

With regard to claim 178, the capillary electrophoresis device in Figure 3 of Klein et al. exposes the sensor (i.e. the DNA of Agilent) to an electric field.

With regard to claim 179-180, Figure 8 of Klein et al. illustrates a data acquisition system that inputs data relating to properties of the electric field. Figure 8 of Klein et al. also illustrates a power supply (label 670) for controlling power to the system.

With regard to claims 181-183, Figures 6 and 8 of Klein et al. illustrate use of a computer program product to control delivery through a channel of a substrate. Page 5 of Agilent further illustrates delivery of a plurality of agents to the sensors in a system such as in Figures 6 and 8 of Klein et al. The cover Figure of Klein et al. illustrates a plurality of reservoirs to apply to cell based biosensors.

With regard to claims 185-188, Figure 8 of Klein et al., label 510 shows use of a vacuum pump while the eighth page of Agilent illustrates a positive pressure profile within a capillary substrate. Figure 1 of Jardemark et al. illustrates a microfluidic chip that contains samples and buffers in communication with a microchannel in communication with a waste reservoir.

With regard to claims 189, Figure 3 of Klein et al. illustrates a fluid delivery mechanism delivering two or more substantially separate fluid streams through a microchannel (i.e. see pages 4-5 and Agilent for examples of such separate fluid streams).

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al. and Agilent by use of the patch clamp capillary electrophoresis apparatus of Jardemark et al. wherein the motivation would have been that this patch clamp method on cells is useful in fractionating biological tissues and can sense very low concentration and small-sized analytes (see penultimate paragraph of the introduction on page 3428 of Jardemark et al.). It would have been further obvious to apply the pressures (positive and negative) and waste collection of Klein et al. to the microfluidic device of Jardemark et al. wherein the motivation would have been that such pressures and waste collections permit more convenient analyses of the cell based biosensors [see Figure 1 and its caption in Jardemark et al.]

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant argues that since the first 35 U.S.C. 103 rejection is allegedly deficient, the instant rejection is also deficient. This argument is not persuasive because the combination of references teaches all of the limitations of the instantly rejected claims. It is noted that the document of Ishmagilov et al. is NOT included in this rejection because the amended limitation of requiring "substantially separate fluid streams" is not in independent claim 162 or any of dependent claims 163-189.

The following rejection is newly applied and is necessitated by amendment:

35 U.S.C. 103 Rejection #6:

Claim 125 is rejected under 35 U.S.C. 103(a) as being unpatentable over Klein et al. in view of the Agilent brochure in view of Ishmagilov et al. as applied to claims 1-3, 8-13, 15-18, 24-33, 36-39, 43-94, 97-98, 100-122, 124-134, 138-139, 142-149, 151, 153-155, and 159-161 above, and further in view of Couderc et al. [Electrophoresis, 1998, volume 19, pages 2777-2790].

Claim 125 is further limiting wherein the light source is a laser in optical communication with a sensor in a microchannel.

Klein et al., Agilent, and Ishmagilov et al. make obvious a capillary electrophoresis system for detecting the presence and properties of sensors and analytes, as discussed above.

Klein et al., Agilent, and Ishmagilov et al. do not teach use of a laser in optical communication with a sensor.

The study of Couderc et al. teaches a CE apparatus with a laser induced fluorescence detector (see abstract of Couderc et al.)

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the capillary electrophoresis apparatus of Klein et al. and Agilent and the microfluidic substrate of Ishmagilov et al. by use of the LIF detector in Couderc et al. wherein the motivation would have been that using such a detector allows greater sensitivity and as a result less sensor amount (see introduction on page 2777 of Couderc et al.).

Response to arguments:

Applicant's arguments filed 2 September 2009 have been fully considered but they are not persuasive.

Applicant argues that since the first 35 U.S.C. 103 rejection is allegedly deficient, the instant rejection is also deficient. This argument is not persuasive because the combination of references teaches all of the limitations of the instantly rejected claims.

Conclusion

No claim is allowed.

Papers related to this application may be submitted to Technical Center 1600 by facsimile transmission. Papers should be faxed to Technical Center 1600 via the central PTO Fax Center. The faxing of such pages must conform with the notices published in the Official Gazette, 1096 OG 30 (November 15, 1988), 1156 OG 61 (November 16, 1993), and 1157 OG 94 (December 28, 1993)(See 37 CFR § 1.6(d)). The Central PTO Fax Center Number is (571) 273-8300.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Russell Negin, whose telephone number is (571) 272-1083. The examiner can normally be reached on Monday-Friday from 7am to 4pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's Supervisor, Marjorie Moran, Supervisory Patent Examiner, can be reached at (571) 272-0720.

Information regarding the status of the application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information on the PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

/Russell S. Negin/
Examiner, AU 1631
22 September 2009